

BMES Cell Team 2019-2020: Lab 3 Immunostaining Protocol

*Volumes are for a 24 well plate

Day 1

1. Fix cells
 - a. Add 500uL 4% paraformaldehyde to each well
 - b. Incubate for 15 mins
 - c. Perform in fume hood because PFA is TOXIC!
2. Wash with PBS-Tween
 - a. 500uL per well
 - b. Wash three times for five minutes each
3. Add 500uL of Triton to each well
 - a. Incubate for 5 minutes
4. Wash with PBS-Tween
 - a. 500uL per well
 - b. Wash three times for five minutes each
5. Block with 500uL 5% BSA-PBS-Tween for an hour an hour at RT
6. Add primary antibodies
 - a. Do not add primaries to one well of each sample so they can act as the control
 - b. Rabbit anti-SOX2 diluted added at 1:200
 - c. Add 500uL to each well x 10 wells = 5mL
 - i. Will not make extra because antibodies are expensive
 - ii. 5mL PBS-Tween + 25uL Rabbit anti-SOX2
 - d. Incubate at 4C overnight

Day 2

1. Wash with PBS-Tween
 - a. 500uL per well
 - b. Wash three times for five minutes each
2. Add secondary antibodies
 - a. When making, cover in foil to avoid photobleaching
 - b. Three secondaries
 - i. Hoescht -- stains nuclei (stains all cells)
 - ii. Goat anti-Rabbit, -- binds to Rabbit anti-SOX2
 - iii. Phalloidin -- binds to actin, already fluorescent
 - iv. Add Hoescht and Goat anti-Rabbit to all wells, add Phalloidin only to wells with primaries
 - c. Wells without primary (control wells)
 - i. Add 500uL to each well x 2 wells = 1mL
 - ii. 1mL PBS-Tween + 1uL Hoescht + 1uL Goat anti-Rabbit
 - d. Wells with primary
 - i. Add 500uL to each well x 10 wells = 5mL

- ii. 5mL PBS-Tween + 5uL Hoescht + 5uL Goat anti-Rabbit + 10uL Phalloidin
 - e. Incubate at RT covered in foil for one hour
- 3. Use EVOS microscope to image
 - a. Turn off lights to avoid photobleaching
 - b. Look at cells under phase (transmitted) at 4x
 - c. Look at fluorescent channels
 - i. Hoescht -- DAPI
 - ii. SOX2 -- GFP
 - iii. Actin (Phalloidin) -- RFP
 - 1. View at 40x to see different actin cytoskeleton morphology between cell lines